Indirect spectrophotometric determination of captopril in pharmaceutical tablets and spiked environmental samples

Nief Rahman Ahmed
Department of Environmental Technology, Coll of Environmental Technology, University of Mosul, Mosul-Iraq

(NJC)

(Received on 12/9/2012) (Accepted for publication 13/1/2013)

Abstract

A simple, accurate and sensitive indirect spectrophotometric method for the determination of captopril in pharmaceutical preparation (tablets) and environmental water samples has been developed. The method is based on the oxidation of captopril by a known excess of potassium iodate (KIO$_3$) in sulfuric acid medium to form iodide ion which reacts with excess iodate to liberate iodine which then reacts with starch to form a stable blue colored Iodine-starch complex which shows maximum absorbance at 606 nm. Beer’s law was obeyed in the concentration range (2-28 ppm). The molar absorptivity and Sandell’s sensitivity of the colored complex are $1.716 \times 10^4$ l/mol.cm. and 12.66 ng/cm$^2$ respectively. The analytical parameters were optimized and the method was successfully applied to the determination of captopril in pure form, its tablets form and environmental water samples.

Keywords: Captopril, Indirect Spectrophotometry, Pharmaceutical tablets, Environmental Samples
Introduction

Captopril, 1-[(2s)-3-mercapto-2-methyl propionyl]-l-proline, that is used therapeutically as an antihypertensive agent, it acts as a reactive and specific inhibitor of the zinc containing angiotensin converting enzyme, which is also prescribed for congestive heart failure \(^{(1-3)}\).

![Chemical Structure of Captopril](image)

\[\text{C}_{9}\text{H}_{15}\text{NO}_{3}\text{S} \quad 217.3\]

Several analytical methods have been devised for the determination of captopril. These methods include titrimetric methods \(^{(4-10)}\). The instrumental methods include HPLC \(^{(11)}\), polarography \(^{(12)}\), gas chromatography \(^{(13)}\), voltammetry \(^{(14)}\), flow injection \(^{(15)}\), fluorimetry \(^{(16)}\), and indirect atomic absorption method \(^{(17)}\). These methods are not simple for routine analysis and required expensive or sophisticated instruments. Many spectrophotometric reagents were used for the determination of captopril including chloranilic acid \(^{(18)}\), dithionitro benzoic acid \(^{(19)}\), and silver nitrate \(^{(20)}\). In this paper we report a simple, sensitive, and accurate spectrophotometric method for the determination of captopril in pure form, pharmaceutical formulations and environmental water samples. The method based on the reduction of iodate into iodid which reacts with excess iodate to liberate iodine which then reacts with starch to form a stable blue colored Iodine-starch complex that has a maximum absorption at \(\lambda_{\text{max}} 606\) nm.

Experimental

Apparatus

A Genway 6405 UV-visible spectrophotometer with 1.0 cm quartz cells was used for absorption measurements.

Reagents

All chemicals used were of analytical or pharmaceutical grade and high-purity water was used throughout. Captopril was obtained from (NDI): the state company for drug industries, Mosul-Iraq.

Captopril stock solution (1000ppm): This solution was prepared by dissolving 0.1 gm of captopril in 100 ml distilled water in a volumetric flask.
Captopril standard solution (100 ppm) (4.6x10^4 M): This solution was prepared by diluting 10 ml of stock solution to 100 ml by distilled water in a volumetric flask.

Potassium iodate solution 0.1% (4.6x10^-3 M): This solution was prepared by dissolving 0.1 gm of Potassium iodate (BDH) in 100 ml distilled water in a volumetric flask.

Sulfuric acid 1N: This solution was prepared by diluting 27.8 ml of (36 N sulfuric acid) to 1L by distilled water in a volumetric flask.

Starch solution 1%: This solution was prepared by dissolving 1gm of soluble starch in 50 ml of formamide (the starch dissolved within 1 minute) then completed to 100ml by distilled water in a volumetric flask.

Recommended procedure

A known volume of sample containing 50-700 μg of captopril was transferred into a series of 25ml calibrated flask followed by 1ml of 1N sulfuric acid solution, 3ml of 1% starch solution and 1ml of 0.1% potassium iodate solution, the volume was made up to the mark with distilled water. The absorbance was measured at 606 nm against a reagent blank prepared in the same way but containing no captopril drug.

Assay procedure for tablets

Weigh and powder 10 tablets. Dissolve a quantity of the powdered tablets containing 100mg of captopril in about 100 ml distilled water, mixed well for 20 min and then filtered. The filtrate was made up to 1L with distilled water. Treat 3 ml of this solution as mentioned under recommended procedure.

Procedure for water samples

Distilled and tap water samples (100ml) were spiked with 10 mg of captopril. The spiked water samples were analyzed as desired under recommended procedure.

Results and Discussion

Captopril is a reducing agent owing to the presence of thiol group(-SH) in its structure. Captopril reduces KIO₃ to KI, but it is oxidized to disulfide. KI immediately react with the excess KIO₃, resulting in the formation of iodine which then reacts with starch to form blue colored I₂-starch complex. Which show maximum absorbance at 606 nm Fig 1.
Fig [1] : Absorption spectra of 300µg/25ml captopril-KIO₃-Starch against blank.

The various experimental parameters affecting on the development and stability of the reaction product was optimized by changing each variable in turn while keeping all other variables constant.

**Effect of acids**

Trials were made to determine the drug through oxidation with KIO₃ was observed only in acidic solution. So that different volume of 1N of different acids have been tested for this purpose. 3ml of 1N sulfuric acid give high sensitivity than the other. Table 1. This amount was selected for subsequent work.

<table>
<thead>
<tr>
<th>Acids</th>
<th>1ml</th>
<th>3ml</th>
<th>5ml</th>
<th>7ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>HCl</td>
<td>0.01</td>
<td>0.015</td>
<td>0.01</td>
<td>0.01</td>
</tr>
<tr>
<td>H₂SO₄</td>
<td>0.232</td>
<td>0.256</td>
<td>0.256</td>
<td>0.247</td>
</tr>
<tr>
<td>HNO₃</td>
<td>0.211</td>
<td>0.110</td>
<td>0.054</td>
<td>0.020</td>
</tr>
<tr>
<td>H₃PO₄</td>
<td>0.125</td>
<td>0.213</td>
<td>0.222</td>
<td>0.211</td>
</tr>
</tbody>
</table>

**Effect of starch concentration**

The amount of starch solution for maximum color intensity was examined. 2-10 ml were found enough to develop the color to its full intensity, 3 ml was selected in all subsequent experiments.

**Effect of KIO₃ reagent**

The amount of potassium iodate solution for maximal color intensity was examined. The maximum color intensity was reached at 1-3 ml. However 1ml of 0.1% reagent solution was selected for the subsequent work.

**Effect of reaction time**

The time for complete color formation occurred immediately and remained stable for at least 2 hours.

**Effect of order of addition**

To obtain optimum results the order of addition of reagent should be followed as given under the recommended procedure. Otherwise a loss in color intensity was observed.
**Calibration graph**

Employing the conditions described in the recommended procedure a linear calibration graph of captopril is obtained Fig(2), which shows that Beer’s law is obeyed over the concentration range of (2-28) µg/ml with correlation coefficient of 0.999, intercept of 0.015 and slope of 0.079. The conditional molar absorptivity of the product formed was found to be $1.716 \times 10^4$ L. mol-1.cm-1.

![Calibration curve for the determination of captopril.](image-url)
Accuracy and precision of the proposed method.

To evaluate the accuracy and precision of the method, a pure drug solution was analyzed at three different concentrations, each determination being repeated six times the relative error (%) and relative standard deviation (%) values are summarized in Table (2). From Table (2), it is clear that the relative error was less than 1.6% and the method is found to be precise with RSD value less than 2.0%.

For a better picture of reproducibility, a series of experiments were performed in which the standard drug solution was determined at three different levels each day for six days, with all solutions being prepared a fresh each day, each determination being repeated six times. The day-to-day relative standard deviation values were in the range of 1.3-1.9 % and represent the best appraisal of repeatability of the proposed methods.

<table>
<thead>
<tr>
<th>Captopril taken µg</th>
<th>$E_r$ (%)$^a$</th>
<th>RSD %</th>
</tr>
</thead>
<tbody>
<tr>
<td>100</td>
<td>1.2</td>
<td>1.3</td>
</tr>
<tr>
<td>300</td>
<td>1.3</td>
<td>1.9</td>
</tr>
<tr>
<td>600</td>
<td>1.5</td>
<td>1.5</td>
</tr>
</tbody>
</table>

$^a$: Mean of six determinations

Stoichiometry of reaction

The stoichiometry of the reaction between captopril and $KIO_3$ was investigated using job’s method of continuous variation method of equimolar solution($4.6\times10^{-4}$M), the result obtained show that 2:1 captopril to $KIO_3$ at 606 nm Fig(3), and the suggested reaction and structure of the product might be written as:

$$KIO_3 + 5KI + 6H^+ \rightarrow 3I_2 + 3H_2O$$
$$I_2 + \text{Starch} \rightarrow I_2-\text{Starch} \text{ (Blue complex)}.$$
Effect of interferences:

In order to evaluate the selectivity of the developed method for the analysis of pharmaceutical preparations containing captopril, the effect of presence of several substance that can occur in the real sample was investigated. The excipients studied were: talc, magnesium stearate, lactose, methyl paraben, polyvinyl pirrolidone, gelatin, and microcrystalline cellulose. For this study solutions containing captopril and each one of excipients taken separately in concentrations equal or ten-times greater than of captopril were analyzed under the same condition described under recommended procedure. A level of interference was considered to be acceptable if the error was not higher than ± 3% relative to the expected captopril value. No interferences were observed in the determination of captopril in the presence of the excipients studied.

Analytical application

The proposed method was satisfactorily applied to the determination of captopril in its pharmaceutical formulations (tablets) and water samples. The results of the assay of the pharmaceutical formulations reveals that there is close agreement between the results obtained by the proposed method and the lable claim. Table(3), The results were also compared statistically by student t-test and by the variance ratio F-test with those obtained by official BP method (8) at 95% confidence level. The calculated t- and F- values did not exceed the theoretical values indicating that there was no significant differences between the precision of the proposed and literature method as cited in Table(3), And the results of water samples Table (4) show that the recovery values obtained were close to 100%.
Table(3): Determination of captopril in pharmaceutical formulations (tablets).

<table>
<thead>
<tr>
<th>Pharmaceutical formulations (Captosam tablets (NDI))</th>
<th>Labal amount mg</th>
<th>Found by proposed method *mg</th>
<th>official BP method(8)</th>
<th>t value</th>
<th>F value</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>12.5</td>
<td>12.46</td>
<td>12.80</td>
<td>1.95</td>
<td>2.07</td>
</tr>
<tr>
<td>2</td>
<td>25</td>
<td>24.92</td>
<td>24.95</td>
<td>1.60</td>
<td>1.85</td>
</tr>
<tr>
<td>3</td>
<td>50</td>
<td>50.10</td>
<td>50.20</td>
<td>1.45</td>
<td>1.68</td>
</tr>
</tbody>
</table>

* mean value of six determinations.

T values (n=10, at 95% confidence level tabulated value 2.262).
F values (n1-1 and n2-1 =9, at 95% confidence tabulated value 3.18).

Table(4): Determination of captopril in water samples

<table>
<thead>
<tr>
<th>Water samples</th>
<th>Added µg/ml</th>
<th>Found* µg/ml</th>
<th>Recovery %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tap water</td>
<td>100</td>
<td>99.8</td>
<td>99.8</td>
</tr>
<tr>
<td></td>
<td>300</td>
<td>300.6</td>
<td>100.2</td>
</tr>
<tr>
<td></td>
<td>600</td>
<td>598.9</td>
<td>99.8</td>
</tr>
<tr>
<td>Distilled water</td>
<td>50</td>
<td>50.5</td>
<td>101</td>
</tr>
<tr>
<td></td>
<td>200</td>
<td>199.7</td>
<td>99.85</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td>500</td>
<td>100</td>
</tr>
</tbody>
</table>

* mean value of ten determinations.

The proposed method was compared with other reported spectrophotometric methods and found to be superior , (Table 5).
Table (5): Comparison of the existing spectrophotometric methods with the proposed method for captopril.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Method 1</th>
<th>Method 2</th>
<th>Method 3</th>
<th>Method 4</th>
<th>Method 5</th>
</tr>
</thead>
<tbody>
<tr>
<td>Reference</td>
<td>18</td>
<td>19</td>
<td>20</td>
<td>20</td>
<td>Proposed</td>
</tr>
<tr>
<td>λ Max (nm)</td>
<td>520</td>
<td>412</td>
<td>520</td>
<td>550</td>
<td>606</td>
</tr>
<tr>
<td>Linear range (μg/ml)</td>
<td>40-140</td>
<td>2.17-21.7</td>
<td>2.5-50</td>
<td>0.25-4</td>
<td>2-28</td>
</tr>
<tr>
<td>ε(l/mol.cm)</td>
<td>2.246x10²</td>
<td>1.35x10³</td>
<td>0.783x10⁵</td>
<td>1.4x10⁵</td>
<td>1.716x10⁴</td>
</tr>
<tr>
<td>Relative error</td>
<td>Less than 3.4</td>
<td>Less than 2</td>
<td>Less than 0.3</td>
<td>Less than 1.7</td>
<td>Less than 1.6</td>
</tr>
<tr>
<td>Application</td>
<td>Pharmaceuticals</td>
<td>Pharmaceuticals</td>
<td>Pharmaceuticals</td>
<td>Pharmaceuticals</td>
<td>Pharmaceuticals and water</td>
</tr>
</tbody>
</table>

**Conclusion**

The proposed method was simple, accurate, sensitive and low economical cost. Furthermore, the proposed method doesn't require elaboration of procedures, which are usually associated with chromatographic methods. The proposed method could be applied successfully for determination of captopril in pure form as well as in different dosage forms (tablets) and in water samples.

**Acknowledgments**

The author wishes to express gratitude to his former company[ the state company of drug industries and medical appliance (NDI) Ninavah – Iraq for providing gift samples of captopril standard materials and pharmaceutical preparation (tablets).
References
3- Jankovic.H, Nagy.L, and Pellerito.L; Coordination properties of the ACE inhibitor captopril towards Me₂Sn in aqueous solution, and biological aspects of some dialkyl tin (IV) derivatives of this ligand; Journal of organometallic chemistry, 2003; 668: 129-139.
5- Brashy.A; Titrimetric determination of captopril in dosage form; Acta Pharm Hung, 1995; 65: 91-93.
11- Baldands.E; Determination of thiol drugs in pharmaceutical formulations as their 5-pyridinium derivatives by HPLC with ultra violet detection; Fresenius J. Anal. Chem, 1997; 358: 554-555.
Argenometric assayof captopril inbulk
drug and in tablets:Journal of the
Iranian Chemical Society; ,2004; 1:
106-114. 18-I.M.
21-Vogel.A ;A Text book of
quantitative inorganic analysis;Third
22-I.M. Kolthoff, R.Belcher , V.A.
Stenger. And G. Matsuyama, "
Volumetric Analysis, " Interscience
23-Vogel.A; A Text book of macro
and semi micro qualitative inorganic
analysis ;4th edn,Longman group
limited,London,1974;p.372.
24- Marczenko . Z.
Spectrophotometric determination of
elements ," EllitsHoward,